

Molecular Investigation of Protein and Genes Resistance Mechanisms in *Acinetobacter baumannii* Isolates Recovered from Diverse Clinical Specimens in AL-Diwaniyah Province

Diyar Khlaif Flaifel

Department of Biology, College of Sciences, University of AL-Qadisiyah, IRAQ.

Corresponding Author: Diyar Khlaif Flaifel



www.jrasb.com || Vol. 2 No. 3 (2023): June Issue

Received: 05-05-2023

Accepted: 08-05-2023

ABSTRACT

The Search included 300 different clinical specimens from patients at Al-Diwaniyah General Teaching Hospital were collected for the study From September to December 2022. These specimens comprised blood, urine, burns, and wounds. 40(13.3%) *A.baumannii* out of 300 isolates. The focus of the study is to characterize the CarO protein, Int-2 and bla_{oxa-51} resistance mechanisms in *Acinetobacter baumannii* isolates recovered from different specimens. The samples were taken from various clinical specimens, which were then dispersed as follows: 40 isolates were tested for antibiotic susceptibility using the (Antibiotic Susceptibility Test-AST) on 15 (37.5%) swabs from burns, 10 (25%) swabs from wounds, 10 (25%) from urine, and 5 (12.5%) from blood samples. All isolates were found to be antibiotic-resistant, with the exception of polymyxin B, which had a 30% sensitivity rate and a 70% resistance rate. The gene CarO was found in 10 (25%) of the 40 isolates genetically analyzed using the PCR method, the gene Int-2 was found in 8 (20%) of the 40 specimens, and the gene bla_{OXA-51} was found in 19 (47.5%) of the 40 isolates. These results show that the gene CarO is present in proteins that encode the enzyme Carbapenemase. The findings of this investigation confirmed previous findings that bacteria have substantial polymyxin B resistance and are resistant to the majority of antibiotics. For the diagnosis of this bacteria, the bla_{OXA-51} gene was regarded as a diagnostic marker (*A baumannii*). In Conclusion Polymerase chain reaction technique was found to be simple and useful tool for detection of outer membrane proteins *carO*. Class 2-Integron was found to be carried by *A. baumannii* isolates and antibiotic resistance genes were distributed on this integron.

Keywords- bla_{oxa-51} gene, CarO- protein, AST, ESKAPE, I.C.P, Int-2, *Acinetobacter baumannii*.

I. INTRODUCTION

Acinetobacter baumannii have characteristics that distinguish them, namely, they are aerobic, immobile, polymorphic, bacillus, opportunistic and Gram-negative. Immunocompromised individuals, particularly those who spend a lot of time in hospitals, are susceptible to it and it spreads among them. It typically colonizes illnesses of the skin, lungs, and oropharynx. Due to its widespread antibiotic resistance, it is regarded as a red alert bacteria. *Acinetobacter baumannii* causes R.I, pneumonia and UTI infections. Since they can be found in almost all samples of soil and surface water, the *Acinetobacter* genus of organisms is frequently recognized as being widespread in nature. As

said by Shirin (2018), this species is often produced from the sputum or respiratory secretions, wounds, and urine of hospitalized patients. *Acinetobacter* commonly colonizes irrigation solutions and intravenous solutions in a hospital setting (Tiwari *et al.*, 2015). These bacteria are considered a multi-dangerous enemy and have the potential to acquire antibiotic resistance because they have the ability to acquire drug-resistant genes and the ability to form a biofilm that enables them to resist treatments. Most of the resistance genes identified in the *Acinetobacter* strain were recently obtained from bacteria of the genera *Pseudomonas*, *Salmonella*, or *Escherichia*. This was verified by sequence homology and phylogenetic studies (Howard *et al.*, 2012). Immunocompromised people frequently contract *A.*

Baumannii, especially if they've had a lengthy (>90 day) hospital stay. Numerous infectious illnesses, including pneumonia, bloodstream infections (bacteremia and sepsis), meningitis, necrotizing fasciitis, and urinary tract infections, can be brought on by *Acinetobacter* (Castro et al., 2015). In 2017, according to the Centers for Disease Control and Prevention, carbapenem-resistant *Acinetobacter* is thought to have been the cause of 700 estimated fatalities and 8,500 hospitalized patient infections in the United States. Increasing resistance trends were seen for the antibiotics ciprofloxacin (24% vs. 26%), cefepime (14% vs. 16%), cefepime (9% vs. 10%), ceftazidime (18% vs. 20%), piperacillin-tazobactam (15% vs. 21%), and imipenem (9% vs. 10%). (Chua & Alejandria, 2008).

One of the ESKAPE bacteria is *Acinetobacter baumannii*. Infections in hospitals are primarily brought on by pathogens everywhere in the world. They are isolates of multiple resistance to treatments, which are considered one of the most serious problems facing humanity because of the inability to find alternative treatments to eliminate them. Current antibiotics used against *Acinetobacter* infections such as Carbapenems causes side effects like nausea, vomiting, diarrhea, and seizure (Choudhary et al., 2017). The efficacy of such antibiotics is also decreasing due to the continuous increase of resistance ability of *A. baumannii*. Hence, there is a need to search new infection fighting medicines to control microbial infections, more

specifically, *Acinetobacter baumannii*'s. This study is conducted to provide additional option to plant-derived medicines. Bermuda grass (*Cynodon dactylon*) had shown antibacterial effects on Gram-positive and Gram-negative bacteria. This implies that this plant is a promising candidate for the formulation of new medicine that can fight *Acinetobacter* infections (Lee et al., 2010).

The study's objective is to identify the function of the genes *bla_{OXA-51}* and *INT-2*, as well as the protein *carO*, in *Acinetobacter baumannii* isolates recovered from various clinical specimens in hospitals.

II. MATERIALS AND METHODOLOGY

Out of 300 specimens obtained from swabs of burns, wounds, blood, and urine, with respective percentages of 37.5%, 25%, 12.5%, and 25%, the research focused on 40 specimens of *A.baumannii*. The findings of an antibiotic sensitivity test for 40 isolates using 8 antibiotics (Amikacin 30 mg, Piperacillin/Tazobactam 10 mg, Cefepime 30 mg, Cefotaxime 30 mg, Ciprofloxacin 5 mg, Polymyxin B 25 mg, Imipenem 10 mg, and Tetracycline 30 mg) were recorded based on CLSI,2022. The following primers were used in this research to identify the target genes in *A. baumannii* isolates as listed in Table (1).

Table 1: Primers used in this study.

Primer Name	Sequence(5' _3')	Product size (bp)	References
<i>BlaOXA51</i>	F5'-TAATGCTTTGATCGGCCTTG -3 '	353	(Turton <i>etal.</i> , 2006)
	R5'-TGGATGCACTTCATCTTGG -3 '		
<i>CarO</i>	F5'- ATGAAAGTATTACGTGTTTTAGTGACAAC 3	730	(Zhu,2019)
	R5'-TTACCAGTAGAATTCTACACCAACT -3'		
<i>Int-2</i>	F5'-CACGGATATGCGACAAAAAGGT-3'	320	Peymani <i>etal.</i> , 2012)
	R5'-GTAGCAAACGAGTGACGAAATG-3'		

III. RESULTS AND DISCUSSIONS

Acinetobacter baumannii Recognition and Isolation:

Using biochemical testing and visual characteristics, isolates were identified. To isolate *A. baumannii*, a total of 40 samples from urine, swab of burns, and swab of wounds from patients at several hospitals in AL-Diwaniyah City were gathered. The attending physicians and clinical microbiologists decided what was clinically relevant. The samples were first

infected on MacConkey agar and chromagar, and they were then incubated for 24 hours at 37 °C. (300) clinical samples from individuals with various illnesses were used in the current investigation. Out of 300 samples, 40 isolates (or 13.3%) were identified as *A. baumannii*. (The samples consisted of 10 wounds, 15 burns, 10 pee, and 5 blood.) However, 30 of the 300 isolates (10%) were classified as different bacterial species. While there has been no growth (10%).

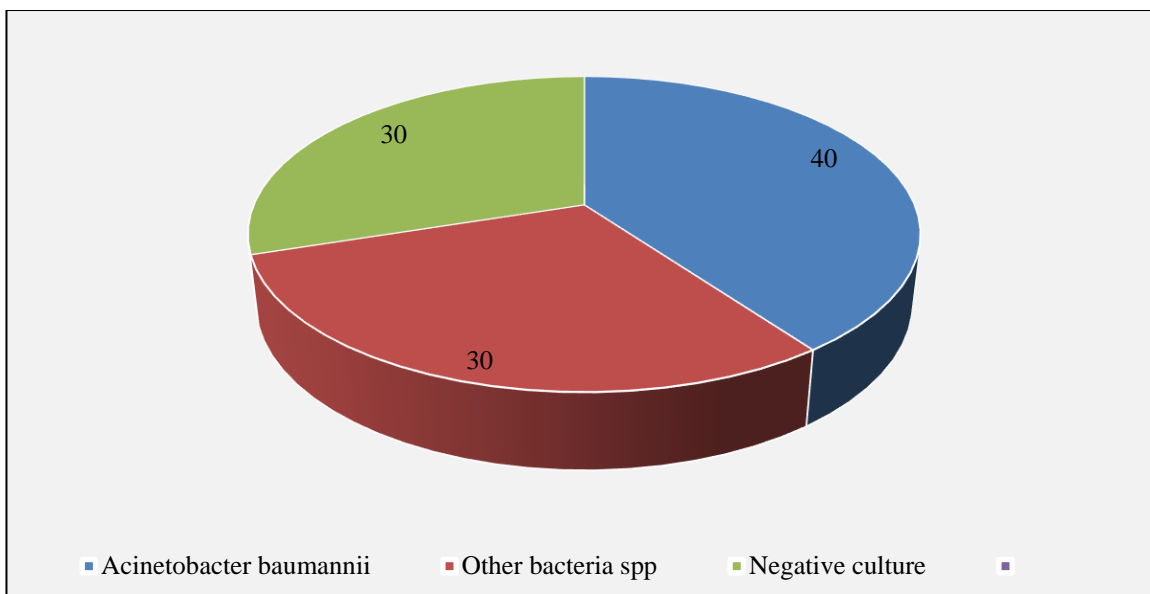


Figure 1: Percentage of Isolation rate in *A.baumannii* from Different Clinical specimens.

In biochemical identification, each isolates gave negative results for indole production, oxidase negative, gas negative, non fastidious, hemolysis negative, nitrate reduction, Vogus Proskaur, motility and urease test while the positive result appear in catalase, simmons' citrate and the results in Kliglar Iron Agar (KIA) developed an alkaline slant, no change bottom, H₂S negative without gas production. All isolated of *A. baumannii* have ability to grow at 37 °C.

Distribution of Acinetobacter baumannii isolates Among Clinical Specimens:

Acinetobacter baumannii isolates had been mended with various percentages from medical specimens, as shown in table (2). Distribution of *A. baumannii* isolates among clinical specimens .

Table 2: Distribution of *A.baumannii* isolates among clinical specimens.

Type of specimens	No. of specimens	No. of Isolates	Percentage
Burn swab	104	15	37.5%
Wound swab	132	10	25 %
Urine	30	10	25 %
blood	34	5	12.5%
Total	300	40	100%

According to the study's findings, out of 300 samples, 40 (13.3%) isolates were found to be *A. baumannii*, and they were distributed among burn swabs (15.5%), wound swabs (10.5%), urine (10.5%), and blood (5.5%). According to Rahi and Raheem's investigations (Raheem, 2020), The greatest finding from this investigation was the rate of bacteria isolates in Hilla, which was 3.33 percent. The isolation rate of an *A. baumannii* isolate was identified as being 21.5% by Raheem (2020), Al-Harmoosh (2015), Al-Kadmy et al. (2019), Al-Baroody and AL-Ghnimi (2020), and Al-Kadmy et al. (2019). Al-Masoudi (2014) discovered that the isolation rate (15%) of the current study was comparable to that of other studies. The isolation rate of

A. baumannii was reported to be 20 (9.23%) in a study by Al-Zubaibi that was published in 2020; however, studies by Hamza and Hadi (2020) came to the conclusion that the isolation rate was 20%.

AST of bacterial isolates:

Using the Kirby-Bauer method, the antibiotic susceptibility profiles of 40 isolates of *A. baumannii* against eight different types of antibiotics were assessed (DDT). Clinical and Laboratory Standard Institute's disk diffusion method for determining antibiotic susceptibility on Muller Hinton Agar. The resulting reading according to CLSI, 2022. Institute CLSI breakpoints standard value (2022).

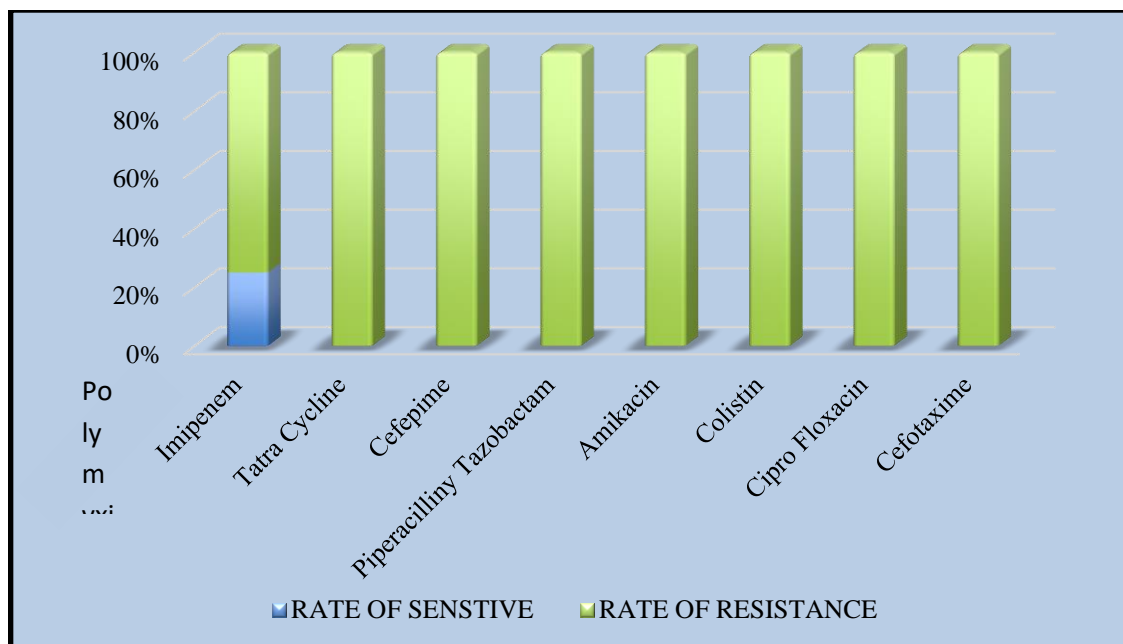


Figure 2: Percentage of Antibiotics Susceptibility Profile of *Acinetobacter baumannii* Isolates Detected by DDT (n=40)

Figure (2), demonstrated that all *A. baumannii* isolates tested in this investigation had the highest level of resistance to most antibiotics. All of the isolates tested positive for resistance to Piperacillin/Tazobactam, which is consistent with research by a local Babylon province study by Al-Warid (2014). Research on the carbapenems group, Imipenem, revealed resistance rates of 100% in 15 isolates, which varied between hospitals in Thailand. Thirapanmethee et al. (2020) discovered that *A. baumannii* isolates were resistant, in contrast to Rahi (2021) who discovered (100%) imipenem resistance rate, and Mshachal et al. (2017), who revealed (50%) polymyxin B resistance rate, sensitive 30%, and resistant 70% in AST. .

Detection of bla_{OXA-51}-like Intrinsic Carbapenemase Gene:

The result of the polymerase chain reaction (PCR) used to detect the bla_{OXA-51} gene as in Figure(3): included 353 bp (Agarose 1%, for 10 minutes at 100 volts, and then decreased to 70 volts, for 60 minutes). visualized under ultraviolet lighting following ethidium bromide staining. Lane L: DNA ladder (100–1500 bp). Lanes 1–15 and 21, 26, 34, 36 indicated bacterial DNA isolates with positive results. Lanes 16–20, 37, 38, 39, and 22–25 represented Negative control is represented by results lane N. the results for 40 *A. baumannii* isolates 19(85%) , Compared with another study conducted in Baghdad, (Abdul-Hussein et al., 2019), recorded that bla_{OXA-51} was detected in 45 (73.77%) isolates among 61 carbapenem -resistant *A. baumannii* isolates. and studies by Al-Hindawi, (2018),bla_{OXA-51} was detected in *A. baumannii* isolates in AL-Diwaniyah hospitals of(100%)and comparable observations were made and other studies by (Al-

Hasnawy ,2018),), recorded that bla_{OXA-51} was detected in 13 (13%) *A. baumannii* isolate. Also studies inTailand by (Thirapanmethee et al .,2020), was detected in all clinical isolates 183. While in (Al-Masoudi,2015) , bla_{OXA-51} was detected in bacterial isolates in percentage (80%) (twelve isolates out of fifteen isolates of bacteria and another study by (Al – Baroody,2020), recorded that bla_{OXA-51} was detected in all isolates of *A. baumannii* 15 isolates (100%). But another study by(Mekkey et al ., 2020) showed the presence of the bla_{OXA-51}-like gene 33 (66%) in out of 50 isolates and, a study by (Anane et al ., 2020), was detected (15%) bla_{OXA-51} gene in *A.baumannii* isolates and study by (Rao et al.,2020), recorded that the bla_{OXA-51} was detected in all isolates of 13 isolates (100%). Further more bla_{OXA-51} was considered a major factor in the resistance to carbapenem in *A.baumannii* (Al-Harmoosh, 2015), and another studies by (Mohamed et al ., 2020) the results revealed that all isolates carried bla_{OXA-51} gene was detected in 82.3% (14/17) in Eygpt . In present results for detection of bla_{OXA-51} gene by molecular methods such as PCR is a gold standard method for confirmation of *A.baumannii* isolates

Detection of INT-2 Gene by polymerase chain reaction:

The PCR amplification of integron genes showed in figure (4). (int-2 gene) showed 320bp. Lanes 38-39and 31,30,29,28) represented positive results of bacterial DNA isolates, Lanes1-25,26,27,28) represented Negative results lane N represent negative control that 8 isolates (20%) harbored class- 2 integron, of. Comparable with another study by (Xu et al ., 2020) , who recorded that no ClassII integron was found in *A. baumannii* which is not greement with the

current study, and another study by (Halaji *et al.*.,2018) , who mentioned that Class II integrons were detected in 78.2% of the bacteriaisolates. And a study by (Salehi *et al.*., 2017), who showed class II integrons that 88.6% of the *A.baumannii* isolates carried the intII genes.And a study by (Zeighami *et al.*., 2019), who recorded that class II (10%) out of 100 bacteria isolates. And a study by (Ardeshiri *et al.*., 2017) , that recorded the class 2 integrons Was 53.8%, out of bacterial isolates. The establishment of MDR *A. baumannii* isolates is significantly influenced by the existence of integrons as a key source of antimicrobial resistance genes within microbial populations.

Detection of CarO protien by polymerase chain reaction:

For molecular detection of outer membrane protein (OMPs) . 40 isolates were analyzed and found in figure (5). (*CarO*gene) showed 730bp. that 10 out of 40 were positive for *carO* gene (25%) , in comparison with study of Zhu *et al.*, (2019) who presented that all isolates were observed were carrying *carO* gene.With the increased number of resistant *Acinetobacter baumannii* isolates.,Furthermore Catel-ferreira at el., (2011), determined the The first proof that the specificities of *carO* channels depend on their fundamental structure and that they have an imipenem (but not meropenem) binding site. Thus, any reduction in *carO* expression would lessen *A. baumannii*'s resistance to this antibiotic. Also, Uppalapati at el .,(2020) proved that decreased expression of the OMPs was significantly associated with carbapenem resistance. The collection of outer membrane proteins (OMPs) is one such arm of *A. baumannii*. When it comes to aiding bacterial adaptation to antibiotic- and host-induced stressors, OMPs in *A. baumannii* play unique functions. OMPs are important allergenic proteins that give bacteria host-fitness benefits such degranulation, stress tolerant, and antibiotic and antibacterial sensitivity. (Uppalapati at el .,2020).

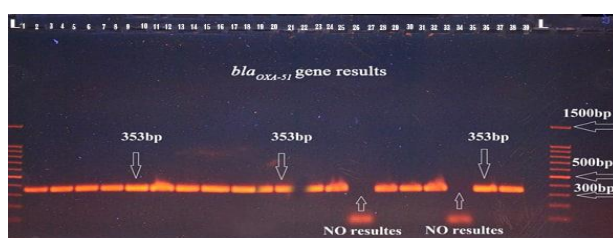


Figure 3: Gel electrophoresis for PCR product (*bla_{OXA-51}* gene) showed 353bp.



Figure 4: Gel electrophoresis for PCR product (*int-2* gene) showed 320bp.

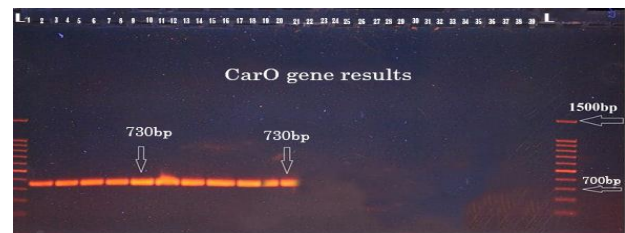


Figure 5: Gel electrophoresis for PCR product (*CarO*gene) showed 730bp.

IV. CONCLUSION

Acinetobacter baumannii is considered a threat to humanity as a result of its ability to acquire resistance to treatments, so it is necessary to seek to find modern treatments that suit the developments of this bacteria, and the gene *bla_{OXA-51}* is considered a diagnostic indicator for bacteria and It is considered *bla_{OXA-51}* gene is essential to identify *Acinetobacter* species as *A. baumannii* .Class 2,Integron was found to be carried by *A. baumannii* isolates and antibiotic resistance genes were distributed on this integron. In the current research, it was discovered that every *A. baumannii* strain was MDR.*A. baumannii* isolates with a high frequency of carbapenemase resistance genes were found in this research.

REFERENCES

[1] Shirin, A.R., Gorji, M., Rezaee, S., Jessop, P.G. and Cunningham, M.F., 2018. CO 2-Switchable-hydrophilicity membrane (CO 2-SHM) triggered by electric potential: faster switching time along with efficient oil/water separation. *Chemical Communications*, 54(61), pp.8478-8481.

[2] Tiwari, A.K., Singh, P.K., Singh, A.K. and De Maio, M., 2016. Estimation of heavy metal contamination in groundwater and development of a heavy metal pollution index by using GIS technique. *Bulletin of environmental contamination and toxicology*, 96(4), pp.508-515.

[3] Howard, A.W., Marcy, G.W., Bryson, S.T., Jenkins, J.M., Rowe, J.F., Batalha, N.M., Borucki, W.J., Koch, D.G., Dunham, E.W., Gautier, T.N. and Van Cleve, J., 2012. Planet occurrence within 0.25 AU of solar-type stars from Kepler. *The Astrophysical Journal Supplement Series*, 201(2), p.15.

[4] Lee, J.Y., Nagano, Y., Taylor, J.P., Lim, K.L. and Yao, T.P., 2010. Disease-causing mutations in parkin impair mitochondrial ubiquitination, aggregation, and HDAC6-dependent mitophagy. *Journal of Cell Biology*, 189(4), pp.671-679.

[5] Castro, M., Expósito-Casas, E., López-Martín, E., Lizasoain, L., Navarro-Asencio, E. and Gaviria, J.L., 2015. Parental involvement on student academic achievement: A meta-analysis. *Educational research review*, 14, pp.33-46.

- [6] Choudhary, M.K., Kataria, J. and Sharma, S., 2018. Evaluation of the kinetic and catalytic properties of biogenically synthesized silver nanoparticles. *Journal of cleaner production*, 198, pp.882-890.
- [7] Al-Masoudi, N.A., Ali, D.S., Saeed, B., Hartmann, R.W., Engel, M., Rashid, S. and Saeed, A., 2014. New CYP17 hydroxylase inhibitors: synthesis, biological evaluation, QSAR, and molecular docking study of new pregnenolone analogs. *Archiv der Pharmazie*, 347(12), pp.896-907.
- [8] Al-Kadmy, I.M., Ibrahim, S.A., Al-Saryi, N., Aziz, S.N., Besinis, A. and Hetta, H.F., 2019. Prevalence of genes involved in colistin resistance in *Acinetobacter baumannii*: first report from Iraq. *Microbial Drug Resistance*, 26(6), pp.616-622.
- [9] AL-Harmoosh, R.A., Jarallah, E.M. and AL-Shamari, A.M., 2015. Coexistence of the blaIMP and blaSIM genes in clinical isolates of *Acinetobacter baumannii* IN Babylon Hospitals-Iraq. *International Journal of PharmTech Research*, 9(7), pp.257-264.
- [10] Hamza, M.M. and Hadi, O.M., 2020. Detection of qnr A and New Delhi metallo-beta-lactamase-1 (blaNDM-1) in *Acinetobacter baumannii* isolated from clinical samples in Hillah hospitals. *Annals of Tropical Medicine and Public Health*, 23, pp.231-401.
- [11] Thirapanmethee, K., Srisiri-A-Nun, T., Houngsaitong, J., Montakantikul, P., Khuntayaporn, P. and Chomnawang, M.T., 2020. Prevalence of OXA-type β -lactamase genes among carbapenem-resistant *Acinetobacter baumannii* clinical isolates in Thailand. *Antibiotics*, 9(12), p.864.
- [12] Mshachal, M.A., Abdulrahman, T.R., Khudair, M.S. and Hassan, J.S., 2017. Molecular detection of multidrug resistant *Acinetobacter baumannii* from different clinical samples. *IRAQI JOURNAL OF MEDICAL SCIENCES*, 15(3).
- [13] Al-Hindawi, F.H. and Ali, A.H., 2018. A pragmatic study of CNN and BBC news headlines covering the Syrian conflict. *Advances in Language and Literary Studies*, 9(3), pp.43-51.
- [14] Mekky, R.H., Thabet, M.M., Rodríguez-Pérez, C., Elnaggar, D.M.Y., Mahrous, E.A., Segura-Carretero, A. and Abdel-Sattar, E., 2020. Comparative metabolite profiling and antioxidant potentials of seeds and sprouts of three Egyptian cultivars of *Vicia faba* L. *Food Research International*, 136, p.109537.
- [15] Anane A, Y., Apalata, T., Vasaiakar, S., Okuthe, G.E. and Songca, S., 2020. Prevalence and molecular analysis of multidrug-resistant *Acinetobacter baumannii* in the extra-hospital environment in Mthatha, South Africa. *Brazilian Journal of Infectious Diseases*, 23, pp.371-380.
- [16] Mohamed, S.A., El-Sakhawy, M. and El-Sakhawy, M.A.M., 2020. Polysaccharides, protein and lipid-based natural edible films in food packaging: A review. *Carbohydrate Polymers*, 238, p.116178.
- [17] Halaji, M., Rezaei, A., Zalipoor, M. and Faghri, J., 2018. Investigation of class I, II, and III integrons among *Acinetobacter Baumannii* isolates from hospitalized patients in Isfahan, Iran. *Oman Medical Journal*, 33(1), p.37.
- [18] Ardeshiri, G., Jamshidi, A. and Keshavarz-Haddad, A., 2017, May. Performance analysis of decode and forward relay network in diffusion based molecular communication. In *2017 Iranian Conference on Electrical Engineering (ICEE)* (pp. 1992-1997).
- [19] Zeighami, H., Valadkhani, F., Shapouri, R., Samadi, E. and Hagh, F., 2019. Virulence characteristics of multidrug resistant biofilm forming *Acinetobacter baumannii* isolated from intensive care unit patients. *BMC infectious diseases*, 19(1), pp.1-9.
- [20] Salehi, S.S.M., Erdogmus, D. and Gholipour, A., 2017, September. Tversky loss function for image segmentation using 3D fully convolutional deep networks. In *International workshop on machine learning in medical imaging* (pp. 379-387). Springer, Cham.
- [21] Zhu, N., Zhang, D., Wang, W., Li, X., Yang, B., Song, J., Zhao, X., Huang, B., Shi, W., Lu, R. and Niu, P., 2020. A novel coronavirus from patients with pneumonia in China, 2019. *New England journal of medicine*.
- [22] Uppalapati, S.R., Sett, A. and Pathania, R., 2020. The outer membrane proteins OmpA, CarO, and OprD of *Acinetobacter baumannii* confer a two-pronged defense in facilitating its success as a potent human pathogen. *Frontiers in microbiology*, 11, p.589234.
- [23] Catel-Ferreira, M., Coadou, G., Molle, V., Mugnier, P., Nordmann, P., Siroy, A., Jouenne, T. and Dé, E., 2011. Structure-function relationships of CarO, the carbapenem resistance-associated outer membrane protein of *Acinetobacter baumannii*. *Journal of antimicrobial chemotherapy*, 66(9), pp.2053-2056.